

## Blood group A is a negative risk factor for peripheric blood stem cell mobilization in allogeneic donors

Tuğçe Nur Yigenoğlu<sup>1</sup>, Mehmet Bakırtaş<sup>1</sup>, Semih Başcı<sup>1\*</sup>, Bahar Uncu Ulu<sup>1</sup>, Derya Şahin<sup>1</sup>, Ali Kılınç<sup>1</sup>, Fatma Nurbüke Şarkışla<sup>2</sup>, Dicle İskender<sup>1</sup>, Nurgül Özcan<sup>3</sup>, Merih Kızıl Çakar<sup>1</sup>, Mehmet Sinan Dal<sup>1</sup>, Fevzi Altuntaş<sup>1</sup>

### Abstract

**Objective:** Many factors, including advanced age and female gender, have been identified as negative factors for peripheric blood hematopoietic stem cell (HSC) mobilization. Similarly, blood group antigens may have an effect on the release of HSCs from the bone marrow niche into the periphery. We aimed to study the effect of ABO and Rh blood groups on peripheral blood HSC mobilization in healthy donors.

**Material and Method:** The data of 314 healthy donors who underwent peripheric blood HSC mobilization in our center were analyzed retrospectively.

**Results:** The number of CD34+ cells collected on the first day and in total was the least in donors with blood group A. A statistically significant relation was found between ABO blood groups and the number of CD34+ cells collected on the first day and in total. No relation was found between Rh positivity and the number of CD34+ cells collected.

**Conclusion:** According to our research in the literature, this is the first study that investigates whether blood groups have an effect on the release of HSCs from the bone marrow niche into the periphery and we observed that blood group A is a negative risk factor for peripheric blood HSC mobilization.

**Keywords:** blood groups, peripheric, stem cell, mobilization, healthy donors

### Introduction

Proteins, glycoproteins, and glycolipids on the surface of erythrocytes define the blood groups' antigens (1). Today, the number of serologically described blood group antigens is more than 600. The majority of blood group antigens are glycoproteins and they are generally described according to their oligosaccharide and amino acid series.

The antigens of ABO blood groups are present on the extracellular membranes of erythrocytes and these antigens are described as complex carbohydrate molecules (2). Besides ABO blood group, Rh blood group is found in the system and at least 45 independent antigens are formed (3).

Progenitor cells constitute only 0.01-0.05% of all nucleated cells in peripheral blood (4,5). Progenitor cells adhere to the micro-environment of the bone marrow with various interactions. The stroma of the bone marrow contains stromal cell-derived factor 1 (SDF-1), vascular cell adhesion molecule (VCAM-1), intercellular adhesion molecule-1 (ICAM-1) and P-selectin glycoprotein ligand,

all of which are ligands for stem cell adhesion molecules (6-8). Inhibition of these receptor-ligand interactions cause increases in the mobilization of progenitor cells (6,7,9).

It has been shown that selectins have a role in the homing of hematopoietic stem cells (HSCs) in the bone marrow (10, 11). On the other hand, ICAM-1 has a role in leukocyte migration, adhesion, and activation, in addition, it may have a role in the regulation of HSC functions (12-16). In previous studies, a relation was found between ABO blood groups and the expression of ICAM-1 and P-selectin (17-21). In the study conducted by Zhang et al, they revealed that blood group A is related to the lowest expression of ICAM-1 and P-selectin (22).

Studies have shown that the number of HSCs infused is closely related to transplant outcomes. Since early neutrophil and platelet engraftment are thought to have a positive effect on survival, risk factors for HSC mobilization should be identified (23-26).

Received 10-05-2019 Accepted 04-06-2020 Available Online 23-06-2020 Published 30-06-2020

1 University of Health Sciences, Dr. Abdurrahman Yurtaslan Oncology Training and Research Hospital, Dept of Hematology and Bone Marrow Transplantation Center, Ankara, TR

2 University of Health Sciences, Dr. Abdurrahman Yurtaslan Oncology Training and Research Hospital, Dept of Internal Medicine, Ankara, TR

3 University of Health Sciences, Dr. Abdurrahman Yurtaslan Oncology Training and Research Hospital, Dept of Clinical Biochemistry, Ankara, TR

\* Corresponding Author: Semih Başcı E-mail: [dr.semihbasci@gmail.com](mailto:dr.semihbasci@gmail.com)



As it has been shown that blood group A is associated with lowest expression levels of ICAM-1 and P-selectin, and as selectins play a role in the bone marrow homing of HSCs and ICAM-1 has a role in cell migration; we hypothesized that blood group A may be a negative risk factor for peripheral HSC mobilization. To reveal this, we aimed to study the effect of blood groups on peripheral blood HSC mobilization in healthy donors.

**Material and Methods**

In this study, the data of healthy allogeneic donors at the age of 18 years and older, who underwent peripheric blood HSC mobilization at our center were analyzed retrospectively. The study was conducted according to the criteria set by the declaration of Helsinki. Each donor signed informed consent before HSC collection. Ethics approval for the study was obtained from the Ethics Committee of the University of Health Sciences, Ankara Dr. Abdurrahman Yurtaslan Oncology Training and Research Center.

Donors were mobilized with a total of 10 µg/kg subcutaneous granulocyte colony-stimulating factor (G-CSF) in 2 divided doses (2 × 5 µg/kg). On the 4th day, if the peripheric blood CD34+ cells' count is ≥50/µL, stem cells started to be collected; if not, G-CSF dose was omitted in the donors whose white blood count (WBC) is ≥ 75.000/mm<sup>3</sup> and the G-CSF dose was reduced by half for the donors whose WBC is ≥50.000/mm<sup>3</sup>, in other donors, G-CSF continued with the same dose and on the 5th day, peripheric blood-derived stem HSCs were collected with a continuous flow blood separator (Fresenius Kabi, COM.TEC, Germany). A total volume of 150 to 400 mL/kg blood was processed for each apheresis at a flow rate of 50 to 60 mL/min. When the number of CD34+ cells collected could not reach the target, G-CSF was continued and the apheresis procedure was repeated the following day. Filgrastim was used in 204 (87.9%) donors and lenograstim was used in 28 (12.1%) donors. Mobilization failure was defined as a CD34+ cell collection below 2 × 10<sup>6</sup> CD34 + cells / kg.

IBM SPSS Statistics (version 21) was used for statistical analysis. Descriptive statistics were used to summarize the data. The suitability of the variables to normal distribution was examined by visual (histogram and graphs) and analytical methods (Shapiro-Wilk and Kolmogorov-Smirnov test). Categorical data were expressed as a ratio, and numerical data were expressed as median and mean ± standard deviation. Kruskal Wallis tests were used for comparison of non-parametric numerical data between groups. P values <0,05 were treated as statistically significant

**Results**

A total of 314 healthy donors were included in the study. The median age was 35 years (range 19 to 65 years). 195 (62,1%) donors were female and 119 (37,9%) were male. The target number of CD34 + cells (5 x 10<sup>6</sup> / kg) was reached on day 4 in 221 (70,4.%) donors. In 213 (67,8 %) donors 1 apheresis procedure was performed to achieve the targeted CD34 + cell count (5 x 10<sup>6</sup> / kg), 2 apheresis procedure was performed in 96 (30,6%) donors and 3 apheresis procedure was performed in 5 (1,6%) donors. There was no donor with the number of HSC collected below 2 x 10<sup>6</sup> / kg. The median number of HSCs in the first and total apheresis products were 6,74 x 10<sup>6</sup> / kg and 7,60 x 10<sup>6</sup> / kg, respectively.

The number of CD34+ cells collected on the first day and in total was the least in donors with blood group A. A statistically significant relation was found between ABO blood groups and the number of CD34+ cells collected on the first day and in total. The median number of the apheresis procedure to achieve target was 1 in all ABO blood groups (Table 1).

277 healthy donors were Rh positive and 37 donors were Rh-negative. No statistically significant relation was found between Rh positivity and the number of CD34+ cells collected on the first day and in total (Table 2).

The number of CD34+ cells collected on the first day and in total was the least in A Rh-positive allogeneic donors (Table 3).

**Table 1:** Mobilization results of donors according to ABO blood groups

Blood Groups	CD34+ cells collected on the 1st day (median) (10 <sup>6</sup> /kg)	CD34+ cells collected in total (median) (10 <sup>6</sup> /kg)	CD34+ cells collected on the first day (median) (/µl)
A (n=113)	5,77 (2-16,6)	6,90 (2,8-20,9)	1073 (200-10207)
B (n=67)	7,93(2,9-26,3)	8 (3-26,3)	1637 (432-5030)
0 (n=114)	6,94 (2 -21)	8 (2,9-21)	1472 (356-4375)
AB (n=20)	7,08 (2,9-16,6)	8,67 (4,7-16,6)	1773 (827-3067)
p-value	0,003**	0,039*	0,001**

**Table 2:** Mobilization results of donors according to Rh groups

Blood Group	CD34+ cells collected on the first day (median) (10 <sup>6</sup> /kg)	CD34+ cells collected in total (median) (10 <sup>6</sup> /kg)	CD34+ cells collected on the first day (median) (/µl)
Rh positive (n=277)	6,83 (2 -26,3)	7,64 (3-26,3)	1317 (200-5030)
Rh negative (n=37)	6,4 (2,5-12,9)	6,9 (2,9-20,9)	1252 (510-10207)
p-value	0,508	0,382	0,547

**Table 3:** Mobilization results of donors according to ABO and Rh groups

Blood Group	CD34+ cells collected on the first day (median) ( $10^6$ /kg)	CD34+ cells collected in total (median) ( $10^6$ /kg)	CD34+ cells collected on the first day (median) ( $\mu$ l)
A Rh (+) (n=95)	5,59 (2-16,6)	6,77 (2,8-16,6)	1065 (200-2708)
A Rh (-) (n=18)	5,71(2,5-11,2)	7(4,5-20,9)	1221 (800-10207)
B Rh (+) (n=62)	7,94 (2-26,3)	8 (2-26,3)	1728 (432-5030)
B Rh (-) (n=5)	7,15(2,9-12,9)	8,5 (3,5-12,9)	1349 (560-1631)
O Rh (+) (n=102)	6,87 (2-21)	8 (2,2-21)	1494 (356-4375)
O Rh (-) (n=12)	8,59 (2,8-11,6)	7,40 (2,9-11,6)	1162 (510-2346)
AB Rh (+) (n=18)	6,69 (2,9-16,6)	8,3 (4,7-16,6)	1773 (827-3067)
AB Rh (-) (n=2)	8,67(8,4-8,8)	8,67 (8,5-8,8)	2085 (1297-2874)
p-value	0,042**	0,193	0,000**

## Discussion

Allogeneic stem cell transplantation (Allo-SCT) is a potentially curative treatment for a variety of benign and malignant hematological diseases. A successful Allo-SCT depends on the infusion of an adequate quantity of HSCs (27,28). Compared to HSC mobilization in healthy donors, there are more factors affecting the amount of HSCs collected in patients with hematologic malignancy, such as underlying disease, radiotherapy history, type, and number of chemotherapies. As there are more factors affecting the amount of HSCs collected in patients with hematologic malignancy, in this retrospective study, we aimed to analyze the effect of donor blood groups on peripheral blood HSC mobilization in healthy donors.

Mobilization failure rate has been reported as 2-40% in various studies (29-34). The prevalence of mobilization failure was 7.6% in the study conducted by Özkurt et al., where mobilization failure was defined as a collection of less than  $2 \times 10^6$  CD34+ cell/kg (35). In our study, similarly, mobilization failure was accepted as a cell collection less than  $2 \times 10^6$  CD34+ cell/kg; and there was no mobilization failure.

To prevent mobilization failure, the factors that have a negative impact on the amount of CD34+ cells collected should be demonstrated. Many factors, including advanced age and female gender, have been identified as negative factors for peripheral blood HSC mobilization (36-38). Similarly, blood group antigens may have an effect on the release of HSCs from the bone marrow niche into the periphery. In a European ancestry population, it was observed that the A1 blood group allele was associated with the lowest expression levels of ICAM-1 and P-selectin (20). In our study, the number of CD34+ cells collected on the first day and in total was the least in donors with blood group A. A statistically significant relation was found between ABO blood groups and number of CD34+ cells collected on the first day and in total.

Human leukocyte antigen (HLA) allele compatibility, young age, and male donor are important factors in allogeneic donor selection. In order to prevent erythrocyte engraftment failure, major and minor ABO blood group incompatibility between patient and donor is avoided.

However, the choice of mobilization method (G-CSF, G-CSF + chemotherapy, or plerixafor) in autologous stem cell collection is made according to the patient's mobilization risk factors. Therefore, identification of all factors affecting HSC mobilization is of great importance.

## Conclusion

According to our research in the literature, this is the first study that investigates whether blood groups have an effect on the release of HSCs from the bone marrow niche into the periphery and we observed that blood group A is a negative risk factor for peripheral blood HSC mobilization. Further studies are needed to reveal all the factors affecting mobilization in order to achieve an adequate number of CD34+ cells.

**Acknowledgement, Funding:** None.

**Author's contributions:** Design of the study: T.N.Y.; Literature search: T.N.Y., M.B.; Material preparation: B.U.U., D.Ş.; Data collection: M.B., F.N.Ş., A.K.; Analysis: S.B., N.Ö.; Preparation of article and revisions: S.B., D.İ., T.N.Y.; Supervision and critical review: M.K.Ç., M.S.D., F.A.

**Conflict of interest:** The authors declare that they have no conflict of interest.

## References

- Nathan DG, Ginsburg D, Orkin SH, Look AT. Nathan and Oski's Hematology of Infancy and Childhood. Sixth Edition, Philadelphia, Saunders. 2003; 1709-56.
- Storry JR, Olsson ML. The ABO blood group system revisited a review and update. Immunohematology. 2009; 25:48-59.
- Schwarz HP, Dorner F. Karl Landsteiner and his major contributions to haematology. Br J Haematol. 2003; 121:556-65.
- Schulz C, von Andrian UH, Massberg S. Hematopoietic stem and progenitor cells: their mobilization and homing to bone marrow and peripheral tissue. Immunol Res. 2009;44 (1-3):160-68.
- Reddy RL. Mobilization and collection of peripheral blood progenitor cells for transplantation. Transfus Apher Sci. 2005;32(1):63-72.
- Bensing W, DiPersio JF, McCarty JM. Improving stem cell mobilization strategies: future directions. Bone Marrow Transplant. 2009;43(3):181-95.

7. Bonig H, Papayannopoulou T. Mobilization of hematopoietic stem/progenitor cells: general principles and molecular mechanisms. *Methods Mol Biol.* 2012; 904:1–14.
8. Liu Y-F, Zhang S-Y, Chen Y-Y, Shi K, Zou B, Liu J, et al. ICAM-1 Deficiency in the Bone Marrow Niche Impairs Quiescence and Repopulation of Hematopoietic Stem Cells. *Stem Cell Reports J.* 2018; Vol. 11:258–73.
9. To LB, Haylock DN, Simmons PJ, Juttner CA. The biology and clinical uses of blood stem cells. *Blood.* 1997;89(7):2233–58.
10. Xia L, McDaniel JM, Yago T, Doeden A, McEver RP. Surface fucosylation of human cord blood cells augments binding to P-selectin and E-selectin and enhances engraftment in bone marrow. *Blood.* 2004; 104:3091–6.
11. Kansas GS: Selectins and their ligands: Current concepts and controversies. *Blood.* 1996; 88: 3259–87.
12. Lawson C, Wolf S. ICAM-1 signaling in endothelial cells: *Pharmacol Rep.* 2009; 61:22–32.
13. Carrasco Y.R, Fleire S.J, Cameron T, Dustin M.L, Batista F.D. LFA-1/ICAM-1 interaction lowers the threshold of B cell activation by facilitating B cell adhesion and synapse formation. *Immunity.* 2004; 20:589–599.
14. Van de Stolpe A and van der Saag PT: Intercellular adhesion molecule-1. *J Mol Med (Berl).* 1996; 74:13–33.
15. Scholer A, Hugues S, Boissonnas A, Fetler L, Amigorena S. Intercellular adhesion molecule-1-dependent stable interactions between T cells and dendritic cells determine CD8+ T cell memory. *Immunity.* 2008; 28:258–70.
16. Peled A, Kollet O, Ponomaryov T, Petit I, Franitz S, Grabovsky V et al. The chemokine SDF-1 activates the integrins LFA-1, VLA-4, and VLA-5 on immature human CD34(+) cells: role in transendothelial/stromal migration and engraftment of NOD/SCID mice. *Blood.* 2000; 95:3289–96.
17. Paré G, Chasman DI, Kellogg M, Zee RY, Rifai N, Badola S et al. Novel association of ABO histo-blood group antigen with soluble ICAM-1: Results of a genome-wide association study of 6,578 women. *PLoS Genet.* 2008; 4:e1000118.
18. Qi L, Cornelis MC, Kraft P, Jensen M, van Dam RM, Sun Q et al. Genetic variants in ABO blood group region, plasma soluble E-selectin levels and risk of type 2 diabetes. *Hum Mol Genet.* 2010; 19:1856–62.
19. Barbalic M, Dupuis J, Dehghan A, Bis JC, Hoogeveen RC, Schnabel RB, et al. Large-scale genomic studies reveal central role of ABO in sP-selectin and sICAM-1 levels. *Hum Mol Genet.* 2010; 19:1863–72.
20. Otto VI, Damoc E, Cueni LN, Schürpf T, Frei R, Ali S et al. N-glycan structures and N-glycosylation sites of mouse soluble intercellular adhesion molecule-1 revealed by MALDI-TOF and FTICR mass spectrometry. *Glycobiology.* 2006; 16:1033–44.
21. Martinez M, Joffraud M, Giraud S, Baisse B, Bernimoulin MP, Schapira M. et al. Regulation of PSGL-1 interactions with L-selectin, P-selectin, and E-selectin: Role of human fucosyltransferase-IV and -VII. *J Biol Chem.* 2005; 280: 5378–90.
22. Zhang W, Xu Q, Zhuang Y, CHEN Y. Novel association of soluble intercellular adhesion molecule 1 and soluble P-selectin with the ABO blood group in a Chinese population. *Experimental and Therapeutic Medicine.* 2016;12: 909–14.
23. Baldomero H, Gratwohl M, Gratwohl A, et al. The EBMT activity survey 2009: trends over the past 5 years. *Bone Marrow Transplant.* 2011;46(4):485–501.
24. Gratwohl A, Baldomero H, Schmid O, et al. Change in stem cell source for hematopoietic stem cell transplantation (HSCT) in Europe: a report of the EBMT activity survey 2003. *Bone Marrow Transplant.* 2005; 36:575–90.
25. Lee SH, Lee MH, Lee JH, et al. Infused CD34+ cell dose predicts long-term survival in acute myelogenous leukemia patients who received allogeneic bone marrow transplantation from matched sibling donors in first complete remission. *Biol Blood Marrow Transplant.* 2005;11(2):122–8.
26. Stiff PJ. Management strategies for the hard-to-mobilize patient. *Bone Marrow Transplantation.* 1999;23:(Suppl. 2)29–33.
27. Takeyama K, Ohto H. PBSC mobilization. *Transfus Apher Sci.* 2004;31(3):233–43.
28. Pusic I, Jiang SY, Landua S, et al. Impact of mobilization and remobilization strategies on achieving sufficient stem cell yields for autologous transplantation. *Biol Blood Marrow Transplant.* 2008;14(9):1045–56.
29. Suzuya H, Watanabe T, Nakagawa R, Watanabe H, Okamoto Y, Onishi T, et al. Factors associated with granulocyte colony-stimulating factor-induced peripheral blood stem cell yield in healthy donors. *Vox Sang.* 2005; 89(4):229–35.
30. Ings SJ, Balsa C, Leverett D, Mackinnon S, Linch DC, Watts MJ, et al. Peripheral blood stem cell yield in 400 normal donors mobilised with granulocyte colony-stimulating factor (G-CSF): impact of age, sex, donor weight and type of G-CSF used. *Br J Haematol.* 2006;134(5):517–25.
31. Martino M, Bonizzoni E, Moscato T, Recchia AG, Fedele R, Gallo GA, et al. Mobilization of hematopoietic stem cells with lenograstim in healthy donors: efficacy and safety analysis according to donor age. *Biol Blood Marrow Transplant.* 2015;21(5):881–8.
32. Anderlini P, Przepiorcka D, Seong C, Smith TL, Huh YO, Lauppe J, et al. Factors affecting mobilization of CD34+ cells in normal donors treated with filgrastim. *Transfusion.* 1997;37(5):507–12.
33. De la Rubia J, Arbona C, de Arriba F, del Canizo C, Brunet S, Zamora C, et al. Analysis of factors associated with low peripheral blood progenitor cell collection in normal donors. *Transfusion.* 2002; 42(1):4–9.
34. Ozkan MC, Sahin F, Saydam G. Peripheral blood stem cell mobilization from healthy donors. *Transfus Apher Sci.* 2015; 53(1):13–6.
35. Özkurt ZN, Batmaz L, Yeğin ZA, İlhan Ç. Factors affecting hematopoietic stem cell mobilization and apheresis in allogeneic donors: The role of iron status. *Transfus Apher Sci.* 2017; 56(3):470–73.
36. Bertani G, Santoleri L, Martino M, et al. Identification of hematopoietic progenitor cell donor characteristics predicting successful mobilization: results of an Italian multicenter study. *Transfusion.* 2014;54(8):2028–33.
37. Motllo C, Sancho JM, Grifols JR, et al. Mobilization and engraftment of peripheral blood stem cells in healthy related donors >55 years old. *Cytotherapy.* 2014;16(3):406–11.
38. Teipel R, Schetelig J, Kramer M, et al. Prediction of hematopoietic stem cell yield after mobilization with granulocyte-colony-stimulating factor in healthy unrelated donors. *Transfusion* 2015;55(12):2855–63..